mRNA Profiling: Body Fluid Identification Using Multiplex Real-Time PCR.

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This invention relates to a ribonucleic acid (RNA) based assay system for body fluid identification, and in particular to a novel, multiplex, parallel assay system based on messenger RNA expressed in human tissue, and to a method for using the same.
### Real-time PCR Primer and Probe Sequences

<table>
<thead>
<tr>
<th>Body Fluid</th>
<th>Gene</th>
<th>Primer and Probe Sequences/dyes</th>
</tr>
</thead>
</table>
| **Housekeeping Gene** | GAPDH | F: 5'- ATG GAA ATC CCA TCA CCA TCT T (Seq ID 1)  
R: 5'- CGC CCC ACT TGA TTT TGG (Seq ID 2)  
P: 5'-NED- CAG GAG CGA GAT CC (Seq ID 3)  
| GNAS   | F: 5'- TGA ACG CCG CAA GTG GAT (Seq ID 4)  
R: 5'- GGC GGT TGG TCT GTG TGT C (Seq ID 5)  
P: 5'-NED- CTT CAA CGA TGT GAC TGC (Seq ID 6)  
| **Blood** | ALAS2 | F: 5'- GCC GAC ACC CTC AGG TCT T (Seq ID 7)  
R: 5'- GAA ACT TAC TGG TGC CTA AGA TGT T (Seq ID 8)  
P: 5'-VIC- AAG CCA CAC AGG AGA C (Seq ID 9)  
| SPTB   | F: 5'- GCC TTT AAT GCC CTG ATA CAC AA (Seq ID 10)  
R: 5'- GAG TCC TTC AGC TTA TCA AAG TCG AT (Seq ID 11)  
P: 5'-FAM- CAC CGG CCC CAC CT (Seq ID 12)  
| **Saliva** | HTN3 | F: 5'- CTT GGC TCT CAT GCT TTC CAT (Seq ID 13)  
R: 5'- TTT ATA CCC ATG ATG TCT TGT C (Seq ID 14)  
P: 5'-FAM- ACT GGA GCT GAT TCA C (Seq ID 15)  
| STATH  | F: 5'- TCT TGG CTC TCA TGG TTT CCA (Seq ID 16)  
R: 5'- CCA ATT CTA CGC AAA AAT TTC TCT T (Seq ID 17)  
P: 5'-VIC- ATT GGA GCT GAT TCA TC (Seq ID 18)  
| **Semen** | PRM1 | F: 5'- CAG ATA TTA CCG CCA GAG ACA AAG (Seq ID 19)  
R: 5'- AAT TAG TGT CTT CTA CAT CTC GGT CTA T (Seq ID 20)  
P: 5'-FAM- CAG CAC CTC ATG GCT (Seq ID 21)  
| PRM2   | F: 5'- GCC GCA AAA GAC GCT CC (Seq ID 22)  
R: 5'- GCC CAG GAA GCT TAG TGC C (Seq ID 23)  
P: 5'-VIC- TTC TGC AGC CTC TGC GAT (Seq ID 24)  
| **Menstrual Blood** | MMP-7 | F: 5'- GGG AGG CAT GAG TGA GCT ACA (Seq ID 25)  
R: 5'- TGG CAT TTT TTT CAG TCT AGT CAT A (Seq ID 26)  
P: 5'-FAM- AAC AGG CTC AGG ACT AT (Seq ID 27)  
| MMP-10 | F: 5'- TGG TCA CTT CAG CTC TTT TCC (Seq ID 28)  
R: 5'- AAT GGC AGA ATC AAC AGC ATC TC (Seq ID 29)  
P: 5'-VIC- CAC CTT ACA TAC AGG ATT G (Seq ID 30)  

F=forward primer, R=reverse primer, P=probe

**Figure 1**
<table>
<thead>
<tr>
<th>Real-time PCR primer and probe for body fluid identification tripLEXs</th>
<th>Blood Triplex 1</th>
<th>Blood Triplex 2</th>
<th>Saliva Triplex 1</th>
<th>Saliva Triplex 2</th>
<th>Semen Triplex 1</th>
<th>Semen Triplex 2</th>
<th>Menstrual Blood Triplex 1</th>
<th>Menstrual Blood Triplex 2</th>
</tr>
</thead>
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<td>250 nM</td>
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<td>900 nM</td>
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<tr>
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<td>900 nM</td>
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<td>-</td>
<td>-</td>
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<tr>
<td><strong>STATH-VIC</strong></td>
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<td>250 nM</td>
<td>-</td>
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<td>250 nM</td>
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<tr>
<td><strong>GAPDH F</strong></td>
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<td>900 nM</td>
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<td>900 nM</td>
<td>900 nM</td>
<td>1200 nM</td>
<td>300 nM</td>
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<td>75 nM</td>
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</tr>
<tr>
<td><strong>GNAS F</strong></td>
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<td>900 nM</td>
<td>1200 nM</td>
<td>1200 nM</td>
<td>1200 nM</td>
<td>300 nM</td>
<td>150 nM</td>
</tr>
<tr>
<td><strong>GNAS R</strong></td>
<td>-</td>
<td>900 nM</td>
<td>900 nM</td>
<td>1200 nM</td>
<td>1200 nM</td>
<td>1200 nM</td>
<td>300 nM</td>
<td>150 nM</td>
</tr>
<tr>
<td><strong>GNAS-NED</strong></td>
<td>-</td>
<td>250 nM</td>
<td>250 nM</td>
<td>300 nM</td>
<td>300 nM</td>
<td>300 nM</td>
<td>150 nM</td>
<td>-</td>
</tr>
</tbody>
</table>

Figure 2
MESSENGER RNA PROFILING: BODY FLUID IDENTIFICATION USING MULTIPLEX REAL TIME-POLYMERASE CHAIN REACTION (Q-PCR)

This invention claims the benefit of priority from U.S. Provisional Application Ser. No. 60/612,057 filed Sep. 22, 2004 the content of which is incorporated herein by reference.

FIELD OF THE INVENTION

This invention relates to a ribonucleic acid (RNA) based assay system for body fluid identification, and in particular to a novel, multiplex, parallel assay system based on messenger RNA expressed in human tissue, and to a method for using the same.

SEQUENCE LISTING

Appendix A is a sequence listing of DNA sequences identified in FIG. 1; the content of Appendix A is also submitted on a compact disc and is incorporated herein by reference. Attached hereto is one compact disc containing the following files:

SEQ. 1 Housekeeping gene GAPDH (Seq. ID 1)
SEQ. 2 Housekeeping gene GAPDH (Seq. ID 2)
SEQ. 3 Housekeeping gene GAPDH (Seq. ID 3)
SEQ. 4 Housekeeping gene GNAS (Seq. ID 4)
SEQ. 5 Housekeeping gene GNAS (Seq. ID 5)
SEQ. 6 Housekeeping gene GNAS (Seq. ID 6)
SEQ. 7 Blood ALAS2 (Seq. ID 7)
SEQ. 8 Blood ALAS2 (Seq. ID 8)
SEQ. 9 Blood ALAS2 (Seq. ID 9)
SEQ. 10 Blood SPTB (Seq. ID 10)
SEQ. 11 Blood SPTB (Seq. ID 11)
SEQ. 12 Blood SPTB (Seq. ID 12)
SEQ. 13 Saliva HTN3 (Seq. ID 13)
SEQ. 14 Saliva HTN3 (Seq. ID 14)
SEQ. 15 Saliva HTN3 (Seq. ID 15)
SEQ. 16 Saliva STATH (Seq. ID 16)
SEQ. 17 Saliva STATH (Seq. ID 17)
SEQ. 18 Saliva STATH (Seq. ID 18)
SEQ. 19 Semen PRM1 (Seq. ID 19)
SEQ. 20 Semen PRM1 (Seq. ID 20)
SEQ. 21 Semen PRM1 (Seq. ID 21)
SEQ. 22 Semen PRM2 (Seq. ID 22)
SEQ. 23 Semen PRM2 (Seq. ID 23)
SEQ. 24 Semen PRM2 (Seq. ID 24)
SEQ. 25 Menstrual Blood MMP-7 (Seq. ID 25)
SEQ. 26 Menstrual Blood MMP-7 (Seq. ID 26)
SEQ. 27 Menstrual Blood MMP-7 (Seq. ID 27)
SEQ. 28 Menstrual Blood MMP-10 (Seq. ID 28)
SEQ. 29 Menstrual Blood MMP-10 (Seq. ID 29)
SEQ. 30 Menstrual Blood MMP-10 (Seq. ID 30)

BACKGROUND AND PRIOR ART

Conventional methods of body fluid identification use a variety of labor-intensive, technologically diverse techniques that are performed in a series, not parallel, manner and are costly in terms of time and consumption of sample. It used to be standard practice to perform biochemical, serological, and immunological tests to identify the body fluid(s) comprising a biological stain. Increasingly, however, classical methods for body fluid identification have no confirmatory technique for some frequently encountered body fluids. For example, there is no definitive test for the presence of saliva or vaginal secretions, and urine identification can be problematic. The need exists for a more reliable, efficient assay system to supplant conventional methods for body fluid identification.

Previous research in the development of a ribonucleic acid (RNA) based assay system for the identification of body fluids, included considerations for the use of protein and messenger RNA (mRNA) since both are expressed in a tissue-type specific manner. However, multiplex analysis of complex protein mixtures, such as those present in body fluid stains, requires further developments in the field of proteomics. Whereas, messenger RNA (mRNA), the molecular intermediate between genetic deoxyribonucleic acid (DNA) and expressed protein, is, at present, supported by technologies for massively parallel analysis in the field of genomics.

As reported by B. Alberts, et al. Molecular Biology of the Cell 3rd ed., Garland Publishing Inc., NY, 1994, a pattern of gene expression is produced that is unique to each cell type and is evinced by the presence, as well as, the relative abundance of specific mRNAs. Each cell type, such as, blood monocytes, lymphocytes, ejaculated spermatozoa, epithelial cells lining the oral cavity or epidermal cells, has a unique pattern of gene expression.


As more and more tissue-specific genes (mRNAs) are identified for use in the positive identification of body fluids and tissues of forensic importance, there is an increasing need for a device or assay system that provides simultaneous and semi-automated analysis through a common assay format. The novel, multiplex, parallel assay system of the present invention provides a common assay format and offers many advantages over the conventional analysis procedures for body fluid identification.

SUMMARY OF THE INVENTION

A primary objective of the present invention is to provide facile identification of the tissue components present in a body fluid stain.

A second objective of the present invention is to supplant the battery of serological and biochemical tests currently employed in the forensic serology laboratory.

A third objective of the present invention is to provide a common assay format that provides greater specificity in the identification of body fluids with improved timeliness.

A fourth objective of the present invention is to decrease sample consumption during analysis of stains containing body fluids.
A fifth objective of the present invention is to provide simultaneous and semi-automated analysis through a common assay format. A sixth objective of the present invention is to provide a multiplex analysis of body fluids in an assay format that is compatible with DNA analysis methodologies. A preferred method for identifying the presence or absence of at least four body fluids from a human being in a single or mixed solution, includes obtaining a sample solution consisting of a body fluid from a human being, extracting total ribonucleic acid (RNA) from the sample solution, and then subtracting the value of the BFG from the value of the HSK gene, wherein a positive value for both BFGs would indicate the presence of the BFG in the bodily fluid. The more preferred method for identifying the presence or absence of at least four body fluids from a human being in a single or mixed solution, wherein portions of two BFG’s identifying one particular housekeeping gene is selected from at least one of GAPDH and HSK.

Preferably, the BFG is one or more of the group consisting of ALAS2, SPTB, STATH, HTN3, PRM1, PRM2, MMP-7, and MMP-10.

A preferred kit for use in identifying the presence or absence of a bodily fluid, comprising BFG and HSK specific primers and probes is provided. More preferably the BFG specific primers are constructed from one or more of the group consisting of the ALAS2, SPTB, STATH, HTN3, PRM1, PRM2, MMP-7, and MMP-10 genes. The preferred housekeeping gene is selected from at least one of GAPDH and HSK.

Preferably, the primers for the preferred kit, are one or more sequences selected from the group consisting of 5'-att-gaac-atac-cc-aac-tca-tct-t (Seq ID 1), 5'-cgcc-cc-agc-act-gttc-ttg-g (Seq ID 2), 5'-tga-agg-ctg-ggc-gct (Seq ID 4), 5'-ggc-ggt-tgg-tct-ggt-ttc-g (Seq ID 5), 5'-ggc-gac-agc-ctc-ttc-ttg-g (Seq ID 7), 5'-gaa-act-tac-tgg-ttc-ttt-ttc (Seq ID 10), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 15), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 16), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 17), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 18), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 19), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 20), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 22), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 23), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 25), 5'-ggc-ggt-ttg-tct-ggt-gac (Seq ID 26).
Further objectives and advantages of this invention will be apparent from the following detailed description and example of a presently preferred embodiment.

BRIEF DESCRIPTION OF THE DRAWING

FIG. 1 shows the DNA sequences of the PCR primers for the various body fluid-specific genes and housekeeping genes used in the assay.

FIG. 2 shows the optimum DNA primer and probe concentrations for each gene in each triplex.

DESCRIPTION OF THE PREFERRED EMBODIMENTS

Before explaining the disclosed embodiments of the present invention in detail it is to be understood that the invention is not limited in its application to the details of the particular arrangements shown since the invention is capable of other embodiments. Also, the terminology used herein is for the purpose of description and not of limitation.

The invention provides a multiplex, parallel assay to identify the tissue source for at least four body fluids from a human being. The first step in the development of the invention has been the identification of tissue-specific genes that are expressed in only one tissue. For example, each tissue type is comprised of cells that have a unique transcriptome, or gene expression profile, also known as the messenger ribonucleic acid (mRNA) profile. The collection of genes that are expressed within the constellation of differentiated cells that make up a body fluid is called the multicellular transcriptome.

These genes comprise ubiquitously expressed housekeeping (HSK) genes such as GAPDH and GNAS, which are responsible for cell maintenance functions, and genes that are specifically expressed in certain tissues only. The mRNA molecules are present in different quantities depending on the particular species of mRNA and the cell type, and can be classified as abundant, moderately abundant and rare. Although the present invention is dependent on the identification of tissue-specific mRNA molecules, it is not to be considered a limitation of the invention. As previously discussed, tissue-specific genes are still in the process of being identified.

The present invention provides a body fluid identification system containing real time PCR triplexes that are able to recognize each of the candidate genes.

An example of some of the body fluid-specific genes for each of four body fluids that are the subject of the present invention, are identified in the table below:

<table>
<thead>
<tr>
<th>BODY FLUID-SPECIFIC GENES</th>
<th>BODY FLUIDS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Delta-aminolevulinate synthase 2 (erythroid) (ALAS2)</td>
<td>BLOOD</td>
</tr>
<tr>
<td>Beta-spectrin (SPTB)</td>
<td>BLOOD</td>
</tr>
<tr>
<td>Statherin (STATH)</td>
<td>SALIVA</td>
</tr>
<tr>
<td>Histatin 3 (HTN3)</td>
<td>SALIVA</td>
</tr>
<tr>
<td>Prostatein 1 (PRM1)</td>
<td>SEMEN</td>
</tr>
<tr>
<td>Prostatein 2 (PRM2)</td>
<td>SEMEN</td>
</tr>
<tr>
<td>Matrix metalloproteinase 7 (MMP-7)</td>
<td>MENSENTRUAL</td>
</tr>
<tr>
<td>Matrix metalloproteinase 10 (MMP-10)</td>
<td>BLOOD</td>
</tr>
</tbody>
</table>

Gene-specific unlabeled forward and reverse primers along with fluorescent dye-labeled probes for any two genes identified above and a housekeeping gene, are incorporated into a single, triplexed real-time polymerase chain reaction (qPCR). During amplification, the fluorescent dye from the gene-specific probe is released and produces a fluorescent signal that is detected by the real-time PCR instrument. The resulting fluorescence signals identify the body fluids that are present in the sample as a single or mixed stain. For this invention, the body fluids include, but are not limited to, blood, saliva, semen or menstrual blood.

The following example provides further explanation of the present invention.

EXAMPLE

A physiological stain is prepared by collecting human blood via venipuncture; collecting 50 microliter (µl) aliquots placed onto cotton gauze and dried at room temperature. Freshly ejaculated semen is collected in plastic cups, allowed to dry on cotton swabs at room temperature. Buccal scrapings and saliva samples were obtained on cotton swabs, and dried at room temperature. Vaginal secretions, such as menstrual blood, are also obtained and dried at room temperature. For RNA or DNA isolation a 50 microliter (µl) bloodstain or a single semen, menstrual blood or saliva stained cotton swab is used.

For RNA isolation, total RNA is extracted from blood, saliva, vaginal secretions and semen stains with a denaturing solution, such as, guanidine isothiocyanate-phenol:chloroform and precipitated with isopropanol. Next, the extracted total RNA is treated with an enzyme, deoxyribonuclease I (DNase I), and then reverse-transcribed using random decamers as the first strand primer, producing complementary DNA (cDNA). Finally, the cDNA is amplified using gene-specific primers.

Based on the above extraction and separation techniques, a multiplex reverse-transcription polymerase chain reaction (RT-PCR) assay is developed for the definitive identification of all of the body fluids commonly encountered in forensic casework analysis, namely blood, saliva, semen, and menstrual blood. The triplexes are composed of two body fluid-specific genes and a housekeeping (HSK) gene, GAPDH (glyceraldehyde-3-phosphate dehydrogenase) or GNAS (guanine nucleotide binding protein, alpha stimulating) or others, and have been optimized for the detection of blood, saliva, semen, and menstrual blood as single or mixed stains. The methodology is based on gene expression profiling analysis in which the body fluid-specific genes are identified by detecting the presence of appropriate mRNA species. Gene-specific unlabeled primers and the corresponding probe labeled with a fluorescent dye, such as VIC, FAM or NED, are incorporated into a single multiplexed real-time polymerase chain reaction (qPCR). VIC, FAM and NED are commercially available, trademarked fluorescent dyes.

The primers and probes are custom synthesized commercially according to our specifications. Invitrogen Corporation (Carlsbad, Calif.) synthesized the primers and the labeled probes are from Applied Biosystems (Foster City, Calif.). The unique primer and probe sequences are shown in FIG. 1. For the body fluid identification triplexes, optimal primer and probe concentrations were determined experimentally and are as follows: For the blood triplex (SPTB/ALAS2/GAPDH), the primer concentrations were 900/900/300 nM, respectively, and the probe concentrations were 250/250/75 nM, respectively. For the blood triplex (SPTB/ALAS2/GNAS), the primer concentrations were 900 nM for each
primer and the probe concentrations were 250 nM for each probe. For the saliva triplex (HTN3/STATH/GAPDH), the primer concentrations were 900/900/600 nM, respectively, and the probe concentrations were 250/250/150 nM, respectively. For the saliva triplex (HTN3/STATH/GNAS), the primer concentrations were 900 nM for each primer, and the probe concentrations were 200/250/250 nM for each probe. For the saliva triplex (HTN3/STATH/GAPDH), the optimal primer concentration was 900 nM for each gene, and the probe concentration was 250 nM for each gene. For the semen triplex (PRM1/PRM2/GAPDH), the optimal primer concentrations were 900/900/1200 nM, and the probe concentrations were 250/250/150 nM, respectively. For the semen triplex (PRM1/PRM2/GNAS), the optimal primer concentrations were 900 nM for each primer, and the probe concentrations were 200/250/1250 nM, respectively. For the menstrual blood triplex (MMP-7/MMP-11/GAPDH), the primer concentrations were 900/900/150 nM, respectively, and the probe concentrations were 250/250/300 nM, respectively. For the menstrual blood triplex (MMP-7/MMP-11/GNAS), the optimal primer concentrations were 900/900/1200 nM, and the probe concentrations were 200/250/1250 nM, respectively. For the menstrual blood triplex (MMP-7/MMP-11/GAPDH), the optimal primer concentration was 900 nM for each gene, and the probe concentration was 250 nM for each gene. For the semen triplex (PRM1/PRM2/GAPDH), the optimal primer concentrations were 900/900/1200 nM, and the probe concentrations were 250/250/150 nM, respectively. For the semen triplex (PRM1/PRM2/GNAS), the optimal primer concentrations were 900 nM for each primer, and the probe concentrations were 200/250/1250 nM, respectively. For the menstrual blood triplex (MMP-7/MMP-11/GAPDH), the optimal primer concentrations were 900/900/1200 nM, and the probe concentrations were 200/250/1250 nM, respectively. For the menstrual blood triplex (MMP-7/MMP-11/GNAS), the optimal primer concentrations were 900/900/300 nM, respectively, and the probe concentrations were 200/250/150 nM, respectively.

These optimal primer and probe concentrations are also listed in FIG. 2, however, these concentrations should not limit the invention as other concentrations will also be functional.

As the polymerase chain reaction (PCR) product is amplified, the gene-specific probe is cleaved and the fluorescent dye is released producing a fluorescence signal that is detected by the real-time PCR instrument. The resulting fluorescence signals identify the body fluids that are present in the sample as a single or mixed stain. The resulting fluorescence signal and corresponding Ct value can be used to identify those body fluids present by determining the delta Ct (dCt) values generated using the cycle threshold (Ct) of the housekeeping gene (HSK) and the Ct of each of the body fluid specific genes (BFGs), (Ct HSK-Ct BFG). Ct is the cycle number at which the fluorescent signal passes a pre-determined threshold. Depending on the body fluid-specific genes (BFGs) being tested, two positive dCt values would indicate the presence of a particular body fluid, while two negative dCt values would indicate the absence of that body fluid. Whereas, one positive dCt and one negative dCt would indicate the possible presence of that body fluid and suggest the need for additional testing of the sample. However, this approach is not limited to utilizing fluorescence as a detection method, as other detection methods may be used. Also, data analysis is not limited to utilizing the calculations described above, as other methods of calculation may be used.

Advantages of the mRNA-based approach, compared to conventional biochemical analysis include, but are not limited to, greater specificity, simultaneous and semi-automated analysis through a common assay format, improved timeliness, decreased sample consumption and compatibility with DNA extraction methodologies.

While the invention has been described, disclosed, illustrated and shown in various terms of certain embodiments or modifications which it has presumed in practice, the scope of the invention is not intended to be, nor should it be deemed to be, limited thereby and such other modifications or embodiments as may be suggested by the teachings herein are particularly reserved especially as they fall within the breadth and scope of the claims here appended.

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We claim:

1. A method for identifying the presence of one or more body fluids from a human being in a single or mixed stain the fluids selected from the group consisting of non-menstrual blood, menstrual blood, saliva and semen, comprising the steps of:
   a) obtaining a sample stain comprising a body fluid from a human being;
   b) extracting total ribonucleic acid (RNA) from the sample stain;
   c) treating the total RNA from step b) with an enzyme
   d) initiating a reverse-transcriptase (RT) reaction by treating the total RNA from step c) with random decamers to produce cDNA;
   e) amplifying a pair of body-fluid specific genes (BFGs) selected from the pairs: ALAS2 and SPTB, specific for non-menstrual blood, HTN3 and STATH, specific for saliva, PRM1 and PRM2, specific for semen, and MMP-7 and MMP-10, specific for menstrual blood, wherein said amplifying utilizes two primers and a probe for each BFG;
   f) amplifying a housekeeping gene (HSK) selected from one of GAPDH and GNAS, wherein said amplifying utilizes two primers and a probe for each HSK;
   g) determining the presence of the body fluid in the stain by measuring the cycle threshold (Ct) value of the housekeeping gene (HSK) and the Ct values of both BFGs, and then subtracting the Ct value of each BFG from the Ct value of the HSK gene, wherein a positive dCt (HSK-Ct minus BFG-Ct) value of each BFG indicates the presence of the corresponding body fluid in the stain.

2. The method of claim 1, wherein the total RNA is extracted with a denaturing solution.

3. The method of claim 2, wherein the extracting solution is guanidine isothiocyanate-phenol chloroform.

4. The method of claim 3, wherein the extracted total RNA is precipitated with an organic solvent.

5. The method of claim 4, wherein the organic solvent is isopropanol.

6. The method of claim 1, wherein the enzyme used to treat the extracted total RNA is deoxyribonuclease I (DNase I).
Col. 1, line 16, insert:

-- STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

This subject invention was made with government support under the Federal Bureau of Investigation, federal contract number JFB103287. The government has certain rights in this invention. --

Signed and Sealed this
Eighth Day of March, 2011

David J. Kappos
Director of the United States Patent and Trademark Office